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Photorespiration Rate in Mungbean (*Vigna radiata* (L.) Wilczek) Leaves – A further estimation from the quantum yield at photosystem II–.

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Photorespiration rates (PR) in peeled (PL, without epidermis) and nonpeeled (NL) leaves of mungbean (*Vigna radiata* (L.) Wilczek) were estimated from electron transport situations at photosystem II using the chlorophyll fluorescence quenching diagnosis. The number (k_1) of electron equivalents required to produce 9 mol adenosine 5'-triphosphate (ATP) for fixing 1 mol CO_2 was experimentally determined 4.62. Also the number (k_2) of electrons required for releasing 1 mol CO_2 in photorespiration was estimated 7.44 or 9.49 on the assumption that 14.5 or 18.5 mol ATP was consumed for 1 mol CO_2 releasing, respectively. The simultaneous equations were constructed with the parameters of k_1 , k_2 and related elements to calculate PR, and the mutual relationships in energy balance among PR, gross photosynthetic rate (Pg), total photosynthetic rate (TC, $\text{TC} = \text{PR} + \text{Pg}$) and PR ratio (PR/TC) were discussed. The value of PR, 1.75 or 2.05 $\mu\text{mol m}^{-2} \text{s}^{-1}$, in PL was estimated corresponding to $k_2 = 7.44$ or 9.49 in a light intensity of 300 $\mu\text{mol m}^{-2} \text{s}^{-1}$ PPFD, and PR/TC was 18.2 or 20.6%, respectively. The estimation by chlorophyll fluorescence quenching diagnosis may give a more reasonable value of PR as compared with the PR obtained by the subtraction method ($\text{PR} = \text{Pg}$ measured in 2% $[\text{O}_2]$ air – Pg measured in 2% $[\text{O}_2]$ air).

INTRODUCTION

The photorespiration is considered to have a negative effect on CO_2 intake or biomass production in crops, but this phenomenon is also known to play an important role in protecting the photosynthetic function in plants grown under the stress conditions such as drought and high light intensities. The photorespiration rate (PR) in a leaf is usually estimated by subtracting the photosynthetic rate (Pg) measured in the atmospheric air from that measured under the photorespiration-restricting air condition with a high $[\text{CO}_2]$ and/or low $[\text{O}_2]$. The subtracted value are used for the estimation of photorespiratory characteristics in species and cultivars. However, the value of PR obtained in such conditions is predicted to be far different from the actual value in a leaf placed in the natural atmospheric air. In order to know the exact value of PR, another approach is necessary. We examined here the estimation of PR from the electron transport situation of photosystem II (PSII) using the chlorophyll fluorescence quenching diagnosis.

The situation of electron transport from PSII to the sites of CO_2 assimilation and photorespiration is diagnosed by the chlorophyll fluorescence quenching analysis described by Genty *et al.* (1989), Harbinson *et al.* (1990), Krall *et al.* (1990, 1991a, 1991b), Marco *et al.* (1990) and Schreiber *et al.* (1986, 1998). Tokuda *et al.* (1999)

already tried to calculate PR in mungbean leaves from the electron transport. In this trial, they assumed that 9 and 13.5 mol adenosine 5'-triphosphate (ATP) were consumed to fix and release 1 mol CO₂ in assimilation and photorespiration, respectively. However, the energy consumption in photorespiration may be differently estimated depending on how we define the photorespiratory category in relation to ATP consumption. In our study, based on the further estimation of the energy consumption in photorespiratory category, PR of mungbean leaf was calculated and the photorespiratory characteristics were discussed.

MATERIAL AND METHODS

Leaves of mungbean cultivar, Chinese, grown in fertilized 8-L pots were used as the experimental material. CO₂ exchange rate (CER) in detached leaves with and without epidermis was measured with an assimilation chamber (LD-2, Hansatech, UK) of open system (Tokuda *et al.*, 1999), and the sampled air was monitored with an infrared CO₂ analyzer (LI-6262, LI-COR, USA). A leaf, the abaxial epidermis of which was peeled, is termed here PL, and a nonpeeled leaf is NL. Two leaves, Leaf-A and Leaf-B, were used here; in the former the effect of epidermis peeling (change from NL to PL) on photosynthesis, photorespiration and quantum yield (Φ_e) at photosystem II (PSII) was examined, and in the latter the effect of [O₂] variation from 20% and 2% on these parameter values in PL (PL_{20%} and PL_{2%}) were investigated.

The chlorophyll fluorescence quenching was carried out simultaneously with the measurement of CER in Leaf-A and Leaf-B. The intensity of chlorophyll fluorescence at PSII was monitored with a portable fluorescence measurement system (MFMS-2T, Hansatech, UK). The initial fluorescence (F_o) of the leaf of unphotosynthetic situation was measured using a dark-adapted leaf under a measuring beam (3.2 $\mu\text{mol m}^{-2} \text{s}^{-1}$ PPFD, 4.8 kHz) and then a 2 sec pulse illumination of 1,800 $\mu\text{mol m}^{-2} \text{s}^{-1}$ PPFD was given to determine the maximum fluorescence (F_m). After this, the leaf was redarkened for 2 min, and then a red actinic light (300 $\mu\text{mol m}^{-2} \text{s}^{-1}$ PPFD) was continuously illuminated. Fluorescence (F_s) and CER were simultaneously monitored and fluorescence peaks (F_m') were measured by giving pulses of white saturation light.

Based on measurements of fluorescence, Φ_e was calculated from the equations (1).

$$\Phi_e = (F_m' - F_o) / F_m' \quad \dots\dots\dots (1)$$

The equation (2) represents the relationship between gross photosynthetic rate (Pg) and Φ_e (Genty *et al.*, 1989).

$$\text{Pg} = \Phi_e \times L \times 0.5 \times i / k \quad \dots\dots\dots (2)$$

where L is a light intensity supplied to a leaf; L=300 $\mu\text{mol m}^{-2} \text{s}^{-1}$ PPFD is used here. i is the ratio of the photon absorbed by chlorophyll to the incident photon; i= 0.8 is used here.

Assuming that the photon is evenly distributed to the two photosystems, 0.5 is used in the equation (2). The parameter k is the number of electron equivalents required to reduce

1mol CO₂ and calculated from the equation (3).

$$k = \Phi_e \times L_0 / P_g \quad \dots\dots\dots (3)$$

where $L_0 = 0.5 \times L \times i$.

It may be considered that when photorespiration is performed in leaf, light energy supplied to PSII is shared into two functions of CO₂ assimilation and photorespiration. This relationship is expressed as the equations (4) and (5).

$$L_0 = L_1 + L_2 \quad \dots\dots\dots (4)$$

$$P_g = \Phi_e \times L_1 / k_1 - \Phi_e \times L_2 / k_2 \quad \dots\dots\dots (5)$$

where L_1 and L_2 are light energy shared from L_0 and used for CO₂ fixation and photorespiration, respectively. Similarly, k_1 and k_2 are the number of electrons required for CO₂ assimilation and photorespiration, respectively. The values of $\Phi_e \times L_1 / k_1$ and $\Phi_e \times L_2 / k_2$ indicate the total CO₂ assimilation rate (TC) and PR, respectively.

RESULTS AND DISCUSSION

Table 1 shows the parameter values of P_g , Φ_e , L_0 and k determined in Leaf-A and Leaf-B under the different [CO₂] and [O₂] conditions at leaf temperatures of 35°C and 25°C. As shown here, the difference in P_g between the values in PL and NL represents the effect of epidermis peeling. It may be considered that the stomatal resistance removal by epidermis peeling increases CO₂ intake rate and simultaneously enhances intercellular [CO₂], with a result of the restriction of photorespiration.

Table 1. The values of P_g , Φ_e and k measured in different [O₂] and leaf temperatures.

		Measurement condition				Obtained values		
		[CO ₂] μmol mol ⁻¹	[O ₂] %	Temp.	L_0 μmol m ⁻² s ⁻¹	P_g μmol m ⁻² s ⁻¹	Φ_e	k
Leaf-A	NL	350	20	35	120	4.39	0.487	13.31
	PL	350	20	35	120	9.65	0.419	5.21
Leaf-B	PL _{20%}	350	20	25	120	7.88	0.509	7.75
	PL _{2%}	350	2	25	120	12.50	0.481	4.62

Leaf-A (NL and PL) was used for the epidermis peeling test, and Leaf-B (PL_{20%} and PL_{2%}) was measured in 20% and 2% [O₂] air. $L_0 = L \times 0.5 \times i$; $120 \mu\text{mol m}^{-2} \text{s}^{-1} = 300 \mu\text{mol m}^{-2} \text{s}^{-1} \times 0.5 \times 0.8$. The values of k were calculated from the equation (3).

As shown in Leaf-B, Pg of $7.88 \mu\text{mol m}^{-2} \text{s}^{-1}$ measured in 20% $[\text{O}_2]$ increased to $12.50 \mu\text{mol m}^{-2} \text{s}^{-1}$ in 2% $[\text{O}_2]$. The value of k obtained under the photorespirationless condition (2% $[\text{O}_2]$ air) was 4.62, which seems to be close to the theoretical value ($k=4$).

The determination of k_1 and k_2 values is necessary for the estimation of PR from the equations (4) and (5). Tokuda *et al.* used $k_1=4.62$ and $k_2=6.93$ which were determined on the assumption that 9 mole ATP was required to assimilate 1 mol CO_2 and 13.5 mol ATP was required to release 1 mol CO_2 in photorespiration. But here we have assumed two cases where 14.5 (the case I) or 18.5 mol (the case II) ATP is needed to release 1 mole CO_2 in photorespiration. When 9 mole ATP is required for assimilation and 14.5 mol ATP is for photorespiration, the ratio of $k_1 : k_2$ is assumed to be 9:14.5. Based on this ratio, k_1 and k_2 are calculated 4.62 and 7.44, respectively (the case I in Table 2). In the case II, where the nitrogen metabolic reaction is included in photorespiration category, ATP consumption per 1 mol CO_2 releasing is estimated 18.5; accordingly the ratio of $k_1 : k_2$ is 9:18.5 and thus the values of k_1 and k_2 are 4.62 and 9.49, respectively. By substituting $k_1=4.62$, $k_2=7.44$ or 9.49 and the measured values of Pg, L_0 and Φ_e into the equations (4) and (5), the values of TC, PR, L_1 and L_2 are calculated.

Table 2. The estimated number, k_1 and k_2 , of electron equivalents required for 1 mol CO_2 fixing in assimilation and releasing in photorespiration, respectively.

Case	I	II
k_1	4.62 (9)	4.62 (9)
k_2	7.44 (14.5)	9.49 (18.5)

The numbers between parentheses are ATP mol required in assimilation or photorespiration (fixing or releasing of 1 mol CO_2 , respectively).

TC, PR, photorespiration ratio (PR/TC), L_1 and L_2 are listed in Table 3. The values of TC, PR and PR/TC varied with k_2 , but the difference was not large in each parameter between the cases I and II. For example, the values of PR/TC in NL of Leaf-A were 41.8 and 37.9% in the case I and II, respectively; their difference was about 10%. The PR/TC calculated by Tokuda *et al.* (1999) based on $k_1=4.62$ and $k_2=6.93$ using the same leaf under the same condition was 42.9%, showing not so large difference from the above values.

So far PR was usually calculated by subtracting ($\text{PR}=\text{Pg}$ measured in 2% $[\text{O}_2]$ - Pg measured in 20% $[\text{O}_2]$). In Table 4, PR in Leaf-B calculated by the subtraction method is shown together with the related parameters to compare with the PR estimated from electron transport using the equations (4) and (5) in the case II ($k_1=4.62$ and $k_2=9.49$). The value of PR obtained by the subtraction method (PR_{sub}) was 4.62 ($12.50-7.88=4.62$) $\mu\text{mol m}^{-2} \text{s}^{-1}$, which was significantly larger than that ($\text{PR}=1.75 \mu\text{mol m}^{-2} \text{s}^{-1}$) estimated from electron transport. If CO_2 assimilation ($\text{TC}=12.50 \mu\text{mol m}^{-2} \text{s}^{-1}$) and photorespiration ($\text{PR}=4.62 \mu\text{mol m}^{-2} \text{s}^{-1}$) are performed in a light energy of $120 \mu\text{mol m}^{-2} \text{s}^{-1}$ PPFD supplied to PSII, the recalculated value of Φ_e comes up to 0.846, but the actually measured Φ_e is

Table 3. Predicted values of TC, PR, PR/TC, L_1 and L_2 in the cases of I and II.

		Pg	Φ_e	TC	PR	PR/TC	L_1	L_2	L_0
		$\mu\text{mol m}^{-2}\text{s}^{-1}$		$\mu\text{mol m}^{-2}\text{s}^{-1}$		%	$\mu\text{mol m}^{-2}\text{s}^{-1}$		
Case I	Leaf-A								
	NL	4.39	0.487	7.55	3.16	41.8	71.7	48.3	120
	PL	9.65	0.419	10.12	0.47	4.7	111.7	8.4	120
	Leaf-B								
	PL _{20%}	7.88	0.509	9.93	2.05	20.6	90.1	29.9	120
	PL _{2%}	12.50	0.481	12.50	0	0	120	0	120
Case II	Leaf-A								
	NL	4.39	0.487	7.10	2.71	37.9	67.4	52.6	120
	PL	9.65	0.419	10.05	0.40	4.0	110.9	9.1	120
	Leaf-B								
	PL _{20%}	7.88	0.509	9.63	1.75	18.2	87.4	32.6	120
	PL _{2%}	12.50	0.481	2.50	0	0	120	0	120

In the case I, $k_1=4.62$ and $k_2=7.44$ were used in calculation, and in the case II $k_1=4.62$ and $k_2=9.49$ were used. See Table 1 for the case I and II. The values of TC, PR, L_1 and L_2 were calculated from the equations (4) and (5) by substituting the values of Pg, Φ_e , k_1 , k_2 and L_0 .

Table 4. The parameter values estimated in the case II and calculated by subtraction method.

	Pg	TC	PR	PR/TC	Φ_e	$L_1 \times \Phi_e$	$L_2 \times \Phi_e$	$L_0 \times \Phi_e$
	$\mu\text{mol m}^{-2}\text{s}^{-1}$			%		$\mu\text{mol m}^{-2}\text{s}^{-1}$		
Leaf-B in the case II								
PL _{20%}	7.88	9.63	1.75	18.2	0.509	44.5	16.6	61.6
PL _{2%}	12.50	12.50	0	0	0.481	57.7	0	57.7
PL _{sub}	7.88	12.50	4.62 ^{a)}	37.0	0.846 ^{b)}	57.7 ^{b)}	43.8 ^{c)}	101.5 ^{d)}

PR^{a)}= $4.62 \mu\text{mol m}^{-2}\text{s}^{-1}$ was determined by PR=Pg of PL_{2%}-Pg of PL_{20%}. 57.7^{b)} and 43.8^{c)} were calculated as TC $\times k_1$ and PR $\times k_2$, respectively; $k_1=4.62$ and $k_2=9.49$ were used here. 101.5^{d)} was the sum of 57.7^{b)}+43.8^{c)}. 0.846^{b)} was calculated by $101.5 \mu\text{mol m}^{-2}\text{s}^{-1} / 120 \mu\text{mol m}^{-2}\text{s}^{-1}$.

0.509 here. There is a large difference between both values of Φ_e , which may suggest that PR= $4.62 \mu\text{mol m}^{-2}\text{s}^{-1}$ is a considerably overestimated value.

The photorespiration has been regarded as having a functional role of a safety valve for protecting photosynthetic system under the stress conditions. Therefore it may be predicted that a leaf with high PR/TC is superior in keeping its photosynthetic activity longer. The genetic variation in photorespiratory characteristics may become an important selection point in breeding of plant with longer-lived leaves. For making possible such a selection it is indispensable to know the exact value of PR in leaf. From

the result obtained in our study it may be concluded that more reasonable values of PR and PR/TC are predicted based on the electron transport situation diagnosed by the chlorophyll fluorescence quenching analysis.

The Mehler reaction is known as another function of consuming the chemical energy produced by photosystem. The energy consumption in this reaction is differently estimated with a wide range from a small amount (Padmasree and Raghavendra, 1998) to such a large amount as reaching several tens percent of the total chemical energy produced in photosystem (Osmond and Grace, 1995). Environmental conditions are known to be greatly effective on the Mehler reaction, but we predict that the energy consumption in the Mehler reaction is not so large in many cases. Particularly, in our experiment, the light intensity ($300 \mu\text{mol m}^{-2} \text{s}^{-1}$ PPFD) used was below 50% of the photosynthetically saturating light, under the condition of which the Mehler reaction seems to be almost negligible in chemical energy consumption.

REFERENCES

- Genty, B., J. -M. Briantais and N. Baker 1989 The relationship between the quantum yield of photosynthetic electron transport and quenching of chlorophyll fluorescence. *Biochim. Biophys. Acta.*, **990**: 87–92
- Harbinson, J., B. Genty and N. R. Baker 1990 The relationship between CO_2 assimilation and electron transport in leaves. *Photosynth. Res.*, **25**: 213–224
- Krall, J. P. and G. E. Edwards 1990 Quantum yields of of photosystem II electron transport and carbon dioxide fixation in C_3 plants. *Aust. J. Plant Physiol.*, **17**: 579–588
- Krall, J. P. and G. E. Edwards 1991a Environmental effects on the relationship between the quantum yields of carbon assimilation and in vivo PSII electron transport in maize. *Aust. J. Plant Physiol.*, **18**: 267–278
- Krall, J. P., G. E. Edwards and M. S. B. Ku 1991b Quantum yield of photosystem II and efficiency of CO_2 fixation in *Flaveria* (Asteraceae) species under varying light and CO_2 . *Aust. J. Plant Physiol.*, **18**: 369–383
- Marco G. Di, F. Manes, D. Tricoli and E. Vitale 1990 Fluorescence parameters measured concurrently with net photosynthesis to investigate chloroplastic CO_2 concentration in leaves of *Quercus ilex* L. *J. Plant Physiol.*, **136**: 538–543
- Oliver D. J. 1998 Photorespiration and the C_2 cycle. In "Photosynthesis: A comprehensive treaties" ed. by A. S. Raghavendra, Cambridge Univ. Press, Cambridge, pp. 173–182
- Osmond C. B. and S. C. Grace 1995 Perspectives on photoinhibition and photorespiration in the field: quintessential inefficiencies of the light and dark reactions of photosynthesis? *J. Exp. Bot.*, **46**: 1351–1362
- Padmasree, K. and A. S. Raghavendra 1998 Interaction with respiration and nitrogen metabolism. In "Photosynthesis: A comprehensive treaties" ed. by A. S. Raghavendra, Cambridge Univ. Press, Cambridge, pp. 197–211
- Schreiber, U., U. Schliwa and W. Bilger 1986 Continuous recording of photochemical and non-photochemical chlorophyll fluorescence quenching with a new type of modulation fluorometer. *Photosynth. Res.*, **10**: 51–62
- Schreiber, U., W. Bilger, H. Ilormann and C. Neubauer 1998 Chlorophyll fluorescence as a diagnostic tool: basics and some aspects of practical relevance. In "Photosynthesis: A comprehensive treaties" ed. by A. S. Raghavendra, Cambridge Univ. Press, Cambridge, pp. 320–336
- Tokuda, S., F. Kubota, K. Hirao and K. Saitou 1999 Effects of leaf epidermis peeling on the CO_2 exchange rate and chlorophyll fluorescence quenching, and estimation of photorespiration rate from electron transport in mungbean (*Vigna radiata* (L.) Wilczek) Leaves. *J. Fac., Kyushu Univ.*, **43**: 293–302